



**Smart D-N-Adem-kit**

**for profiling**

**(cat #06142)**

**Instruction manual for  
gDNA normalisation from Swabs**

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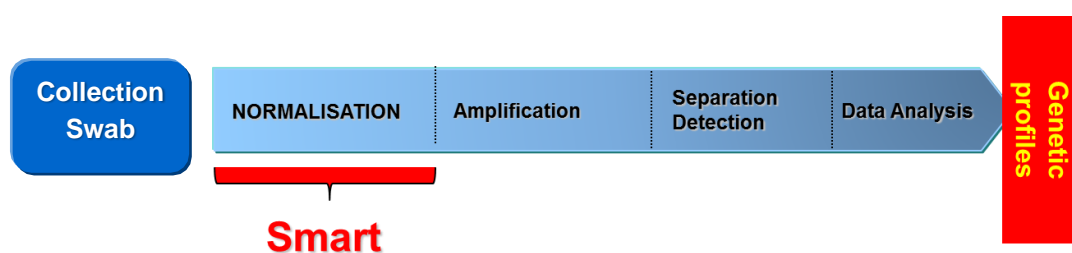
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## Smart D-N-Adem-Kit for profiling/ Swab

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### Introduction

DNA quantities present on swab vary from sample to sample, the collecting devices used, the collection methods applied, the swab-to-FTA™ transfer protocol and also from laboratory to laboratory. Blood and buccal samples often contain substances that can inhibit DNA amplification. Ademtech has developed the Smart D-N-Adem-Kit for profiling for delivering a consistent amount of pure DNA to considerably enhance quality profile and efficiency of forensic laboratories. The DNA is ready to use for STR amplification without any added quantification steps.



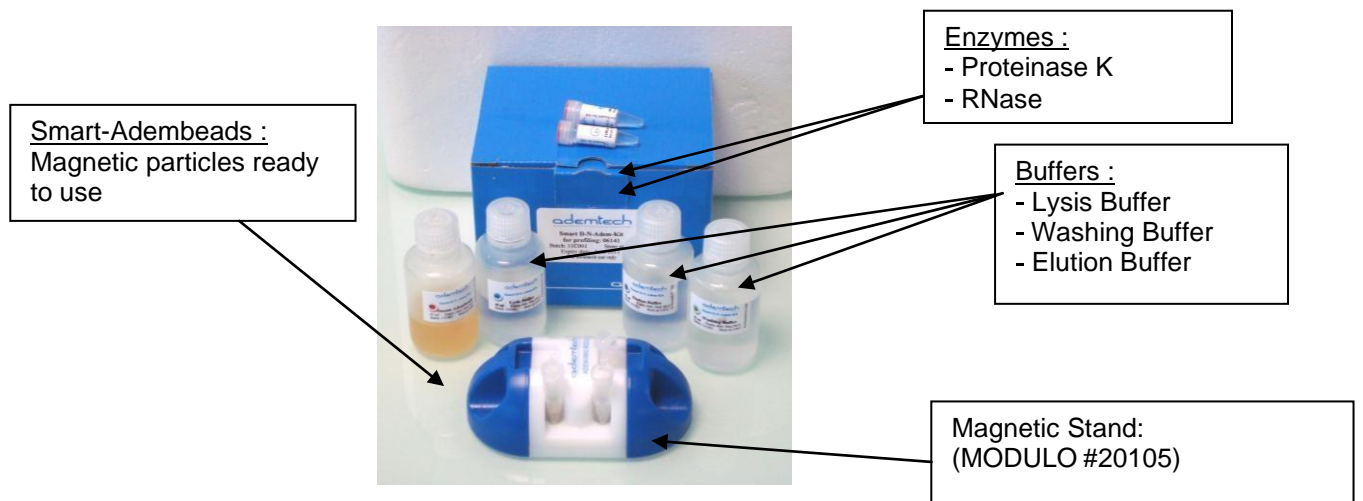
## 1. Smart D-N-Adem-Kit for profiling

### 1.1. Smart-Adembeads Description

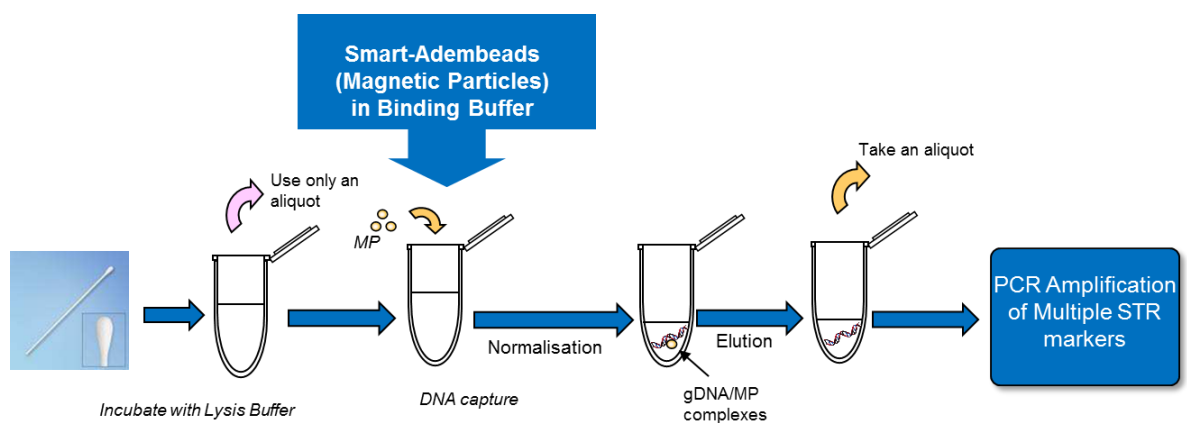
Smart-Adembeads are uniform, monosized beads of 300nm with a large and well defined specific area that ensure optimal reproducibility. Their capacity and performance lead to the capture of a consistent amount of DNA. The beads are composed of a magnetic core encapsulated by a highly cross-linked polymer shell. The high iron oxide content (70%) increases magnetic strength of the beads and ensures rapid magnetic mobility and efficient isolation of nucleic acids. The nanosized beads feature a very low sedimentation rate ideal for fast reaction kinetics, making them particularly suitable for automated assays. Alternative particles from other suppliers often present a random size range distribution, a porous surface associated with an irregular binding capacity; these compromise the reproducibility of your assays.

## 1.2. Smart-D-N-Adem-Kit Description

The Smart D-N-Adem-Kit for profiling contains Smart-Adembeads and specific buffers optimized for capture and normalisation of DNA. Smart-Adembeads offer an innovative surface for gDNA capture and compatible with a direct amplification. The Smart D-N-Adem-Kit procedure allows cleaning gDNA and avoids the use of phenol, ethanol, chloroform and ionic chaotropes that could inhibit PCR.



## 2. DNA normalisation procedure overview



### 3. Kit contents and storage conditions

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**NOTE !** Smart D-N-Adem-Kit avoids the use of harmful organic solvents such as phenol, ethanol, isopropanol or guanidine thiocyanate that can react with acids and bases and generate toxic gas, and eliminates multiple centrifugation steps used in some purification procedures.

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**Kit contents:** Each Smart D-N-Adem-Kit for profiling contains sufficient materials to perform 100 normalisations using the following standard protocol.

**Table 1:** *Materials provided within Smart D-N-Adem-Kit for profiling (# 06142)*

Smart D-N-Adem-Kit (#06142)			
	Amount	Reagents	Storage conditions
<b>R1</b>	50µl	RNase A	+ 2°C to +25°C
<b>R2</b>	250µl	Proteinase K	+ 2°C to +25°C
<b>R3</b>	50ml	Lysis Buffer	+ 2°C to +25°C
<b>R4</b>	10ml	Smart-Adembeads	+ 2°C to +25°C
<b>R5</b>	10ml	Washing Buffer	+ 2°C to +25°C
<b>R6</b>	10ml	Elution Buffer	+ 2°C to +25°C
contains sufficient reagents to perform <b>100 normalisations</b>			

**Storage conditions:** The kits are shipped at room temperature and stored at 4°C or room temperature upon reception.

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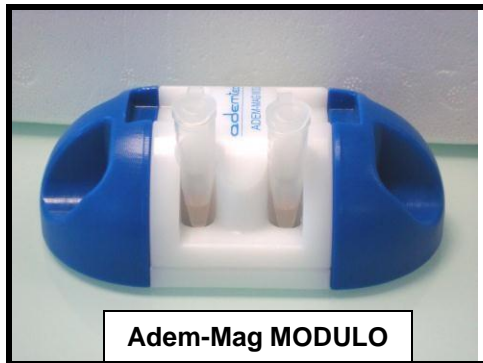
**NOTE !** Properly stored Kits are guaranteed until the expiry date. Note that shipping is realized at room temperature and will not affect stability. All components of the kit have been prepared under nucleases free conditions and have been thoroughly tested to ensure optimal performance.

**IMPORTANT !** Do not freeze the magnetic particles.

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#### 4. Required equipment (not supplied as part of the kit)

- **Adem-Mag MODULO** (#20105, #20108) or **Adem-Mag 96** (#20106):
- Nuclease free microtubes or microplates
- Nuclease free tips
- Disposable gloves
- Thermal shaker or Heat block



### Smart D-N-Adem-kit Protocol

#### 1. Smart-Adembeads Guidelines

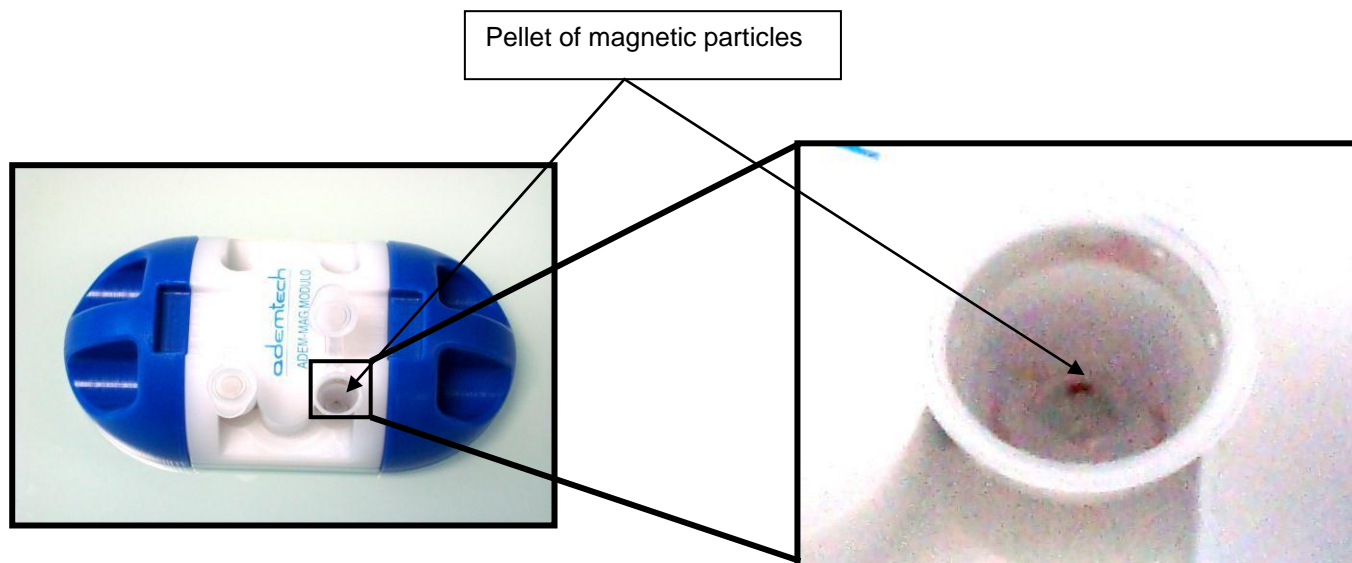
- Before using Smart-Adembeads, thoroughly flick / vortex the bottle to completely resuspend the magnetic particles.
- The Smart-Adembeads are ready to use (there is no need to add Binding Buffer). Colour of the solution is due to the presence of the magnetic particles.

Smart-Adembeads



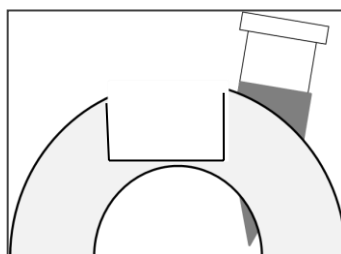
## Smart D-N-Adem-Kit for profiling/ Swab

- During separation steps, let the microtubes containing magnetic particles on the magnet at least 3 minutes. Pay attention to the size of the magnetic pellet which is very small. The magnetic particles pellet is oriented toward the magnet at the back of the microtubes.
- When removing the liquid phase, pipette off carefully, do not aspirate magnetic particles or disturb the magnetic pellet.



## 2. Magnetic Stand Guidelines

- Place magnetic base away from metal objects/magnetic media.
- Insert the microtubes into the sample holder in the correct position.

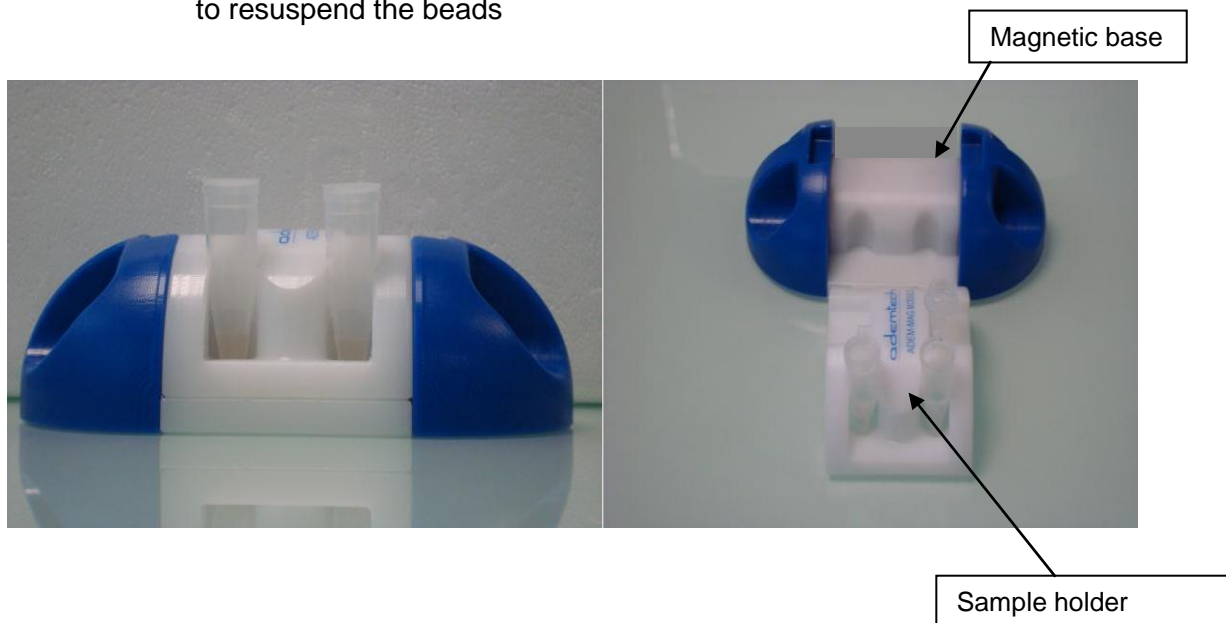


Correct position



### Smart D-N-Adem-Kit for profiling/ Swab

- Insert sample holder into magnetic base. To help optimize magnetic pellet formation ensure that the magnetic stand is correctly assembled before performing washing and elution step.
- The sample holder can be quickly removed from the magnetic base to resuspend the beads



### 3. Smart D-N-Adem-Kit for profiling Protocol

Before starting gDNA extraction procedure, all buffers shall be at room temperature (20-25°C) for optimal performances.

#### **3.1. Prepare samples**

The standard protocol is appropriate for buccals cells on swabs (all types of swabs dedicated to forensic including Omni swab <sup>TM</sup>, Cotton swab).

1. Place the extremity of the dry swab in a 2mL microtube.
2. Break the swab shaft in order to close the tube or push the head of the swab in the bottom of the microtube.

#### **3.2. Perform lysis**

**Preparation of Lysis solution:** Prepare the Lysis solution by combining Lysis Buffer, Proteinase K and RNase in the proportions as indicated below. Mix by pipetting or vortex the tube.

Lysis Buffer	500µl
Proteinase K solution	2,5µl
RNase A solution	0,5µl
<b>Total volume:</b>	<b>503µl</b>

Prepare 503µl of Lysis solution for each swab.

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**IMPORTANT!** RNase must be added in the last to avoid its early degradation by Proteinase K.

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#### **Perform lysis:**

1. Add **500µl of freshly-prepared Lysis solution** to the microtube containing the head of the swab.
2. Mix vigorously for 5 seconds.

## Smart D-N-Adem-Kit for profiling/ Swab

3. Place the tube in a thermal shaker, then incubate at 56°C at 800 rpm for 30 minutes.

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**IMPORTANT!** Temperature can be varied between 50°C and 60°C. You can use a heat block instead of a thermal shaker, however the DNA yield may be lower. For effective recovery, make sure that the sample is immersed by the Lysis Solution during mixing.

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4. After the incubation, mix vigorously for 5 seconds.
5. Use only **50µl of lysat** to perform DNA extraction.

### **3.3. Bind genomic DNA to magnetic particles for normalisation**

1. Distribute **100µl of Smart-Adembeads** in a new microplate or a new microtube.
2. Transfer the lysate (50µl of supernatant) to the Smart-Adembeads.
3. Mix well by pipetting or vortex (during 5s, 1800-2000rpm).
4. Incubate at room temperature for 5 minutes at 900rpm.

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**NOTE!** Increasing the incubation time with the Smart-Adembeads do not affect DNA yield.

Smart-Adembeads are ready to use (there is no need to add Binding Buffer).

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### **3.4. Wash bound DNA**

After binding DNA to the magnetic particles, wash the magnetic particles to remove impurities and inhibitors.

1. Place the tube or microplate on the magnet at least 5 minutes, and discard carefully the supernatant.
2. Remove the tube or microplate from the magnet and add **100µl of Washing Buffer** and mix by pipetting or vortex (during 15s, 900rpm).
3. Place the tube or microplate on the magnet for at least 5 minutes and discard the supernatant.

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**NOTE!** After DNA binding, magnetic particles aggregates could be observed due to the binding of DNA onto the beads. Nevertheless, magnetic particles easily come back in suspension by mixing (pipetting or vortex) during washing step.

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### **3.5. Elute DNA**

After performing the washing step, resuspend the purified DNA and separate the DNA eluate from the magnetic particles.

1. Remove the tube or microplate from the magnet and add **100µl of Elution Buffer**.

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**IMPORTANT!** Do not use water instead of Elution Buffer. Do not dry the magnetic particles

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2. Place the tube or the microplate in a thermal shaker, then mix at room temperature at 900 rpm for 10 minutes (or during 5s, 1800-2000rpm).
3. Incubate overnight at 4°C the tube or the microplate. This step is to maximize DNA recovery.

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**NOTE!** The isolated DNA may be stored at 4°C with the beads for up to one week or at -20°C for longer storage

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**NOTE!** An alternative elution procedure is to heat resuspended particles at 75°C, 900rpm for 5 minutes instead overnight at 4°C.

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4. Place the tube or microplate on the magnet for at least 5 minutes and transfer carefully the liquid (supernatant) which contains the isolated genomic DNA to a new microtube or microplate.

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When removing the liquid phase (supernatant), do not aspirate magnetic particles or disturb the magnetic particles pellet

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## DNA analysis and expected results

For quantifying  
extracted DNA

- We recommend using quantification PCR kit which provides a rapid, sensitive, and accurate method for dsDNA quantification instead of UV absorbance.

- For quantification,
  - Perform Elution (3.5), step 1 through 3
  - Mix by pipetting or vortex and take an aliquot of 1µl or 2µl for a 25µl PCR reaction.

For STR  
analysis

- For STR amplification and analysis, magnetic particles must be removed. Magnetic particles could migrate and interfere with subsequent analysis.
- For STR amplification, we recommended using between 1-2µl of DNA eluate.

## Troubleshooting

Observations	Possible cause	SUGGESTION
<b>Magnetic particles settled in the bottle.</b>	During shipping, magnetic particles settle.	Thoroughly flick / vortex the bottle. Smart-adembeads are stored at 4°C, before using, incubate them at room temperature.
<b>Supernatants contains magnetic particles.</b>	The magnetic stand used is not adapted to the magnetic particles. Incorrect position for microtubes in the sample holder	Keep the tube containing magnetic particles in the magnet for at least 5 minutes
<b>DNA eluate contains magnetic particles</b>	Aggressive pipetting could disturb magnetic pellet	Keep the tube containing magnetic particles in the magnet for at least 5 minutes then pipette out carefully the supernatant Magnetic particles did not perturb DNA quantification but affect STR product migration for analysis
<b>Quantity of DNA eluate is too high for STR analysis</b>	Samples contains lots of cells	Dilutes DNA samples.
<b>Nor or low yield of DNA</b>	Biological sample contains no or low amount of dna	Review protocol steps and reagents additions
	Insufficient amount of magnetic particles added	

## Warranty

This product is only for use in research. The purchaser is responsible to validate the performance of this product for any particular use, and to use the product in compliance with any applicable regulations. The products are warranted to the original purchaser only to conform to the quality and contents stated on the vial and outer labels for duration of the stated shelf life. Ademtech's obligation and the purchaser's exclusive remedy under this warranty is limited either to replacement, at Ademtech's expense, of any products which shall be defective in manufacture, and which shall be returned to Ademtech, transportation prepaid, or at Ademtech's option, refund of the purchase price. Claims for merchandise damaged in transit must be submitted to the carrier.

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